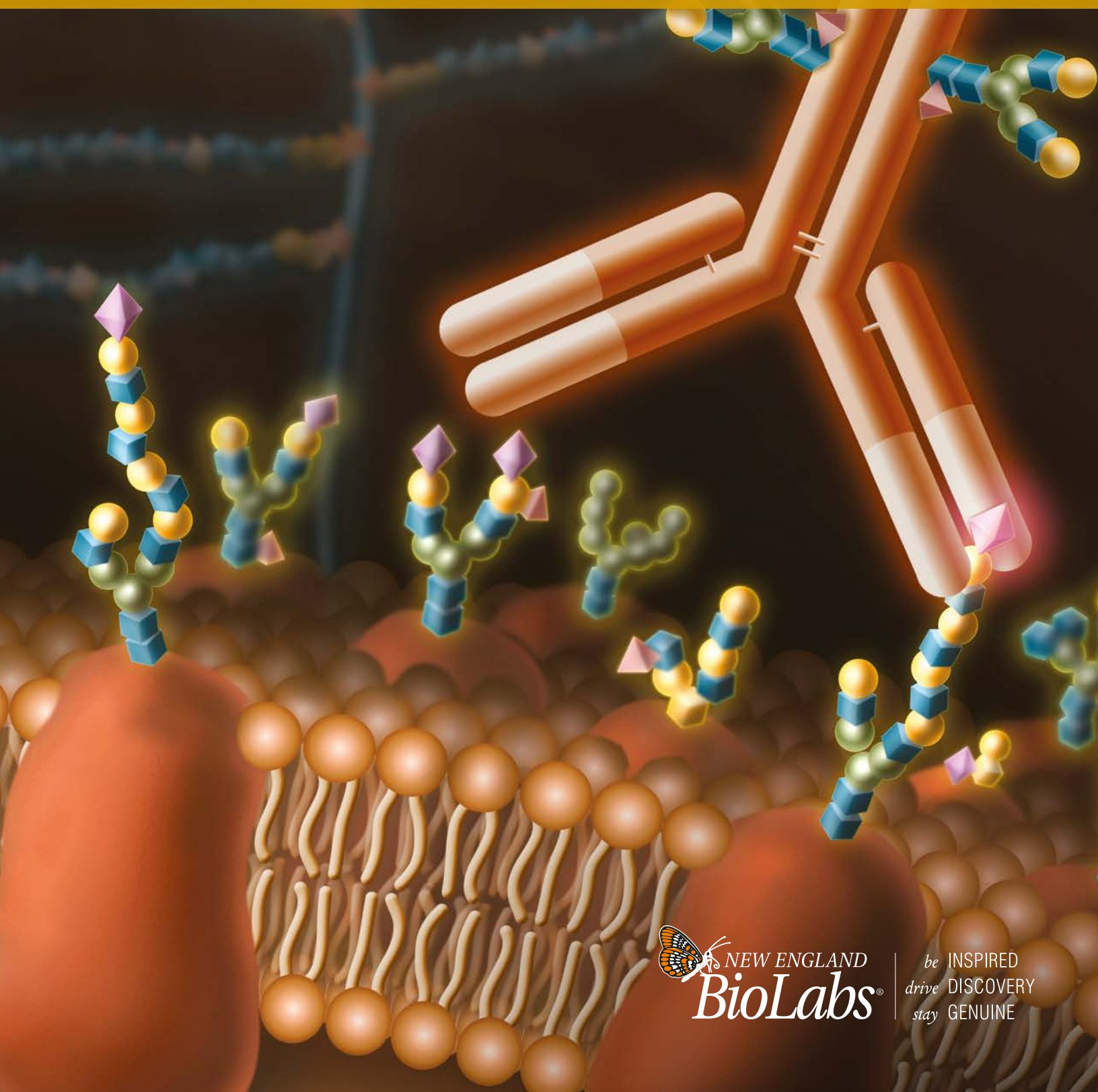


 Now includes O-Glycoprotease

Glycoproteomics

UNDERSTANDING PROTEIN
MODIFICATIONS



 NEW ENGLAND
BioLabs

be INSPIRED
drive DISCOVERY
stay GENUINE



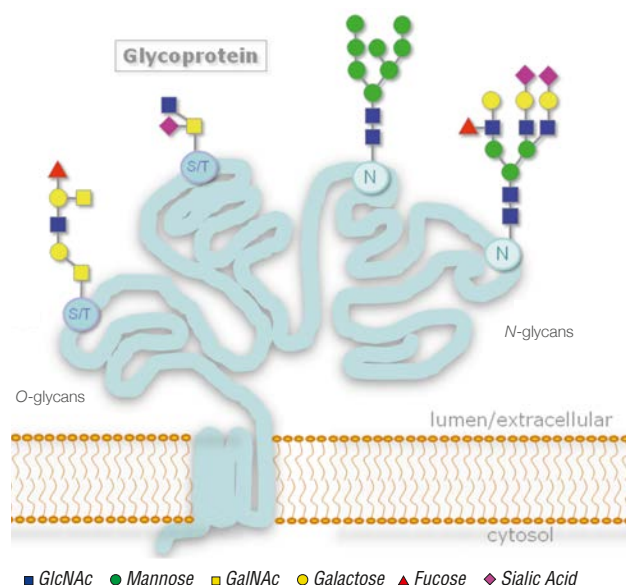
Glycoproteomics Products

New England Biolabs (NEB) offers a selection of endoglycosidases and exoglycosidases for glycobiology research. Many of these reagents are recombinant, and all undergo several quality control assays, enabling us to provide products with lower unit cost, high purity and reduced lot-to-lot variation. All of our glycosidases are tested for contaminants. Since p-nitrophenyl-glycosides are not hydrolyzed by some exoglycosidases, we use only fluorescently-labeled oligosaccharides to screen for contaminating glycosidases.

Glycobiology is the study of the structure, function and biology of carbohydrates, also called glycans, which are widely distributed in nature. It is a small but rapidly growing field in biology, with relevance to biomedicine, biotechnology and basic research. Proteomics, the systematic study of proteins in biological systems, has expanded the knowledge of protein expression, modification, interaction and function. However, in eukaryotic cells the majority of proteins are post-translationally modified (1). A common post-translational modification, essential for cell viability, is the attachment of glycans, shown in Figure 1. Glycosylation defines the adhesive properties of glycoconjugates and it is largely through glycan-protein interactions that cell-cell and cell-pathogen contacts occur, a fact that accentuates the importance of glycobiology. Glycomics, the study of glycan expression in biological systems, relies on effective enzymatic and analytical techniques for correlation of glycan structure with function.

NEB offers a suite of endoglycosidases and exoglycosidases to study glycosylation modifications. Visit www.NEBglycosidase.com for the latest list of enzymes and reagents available from NEB.

Figure 1: N- and O-Glycosylation



N-linked glycosylation occurs through the asparagine residues of the protein, while O-linked glycosylation occurs through serine or threonine.

DOWNLOAD THE NEB AR APP*



Find an overview of glycobiology.



*see back cover for details

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19 Ordering Information

Reference: 1. Spiro, R.G. (2002) *Glycobiology* 12, 43R–56R.



Deglycosylation Enzymes

Several classes of glycans exist, including *N*-linked glycans, *O*-linked glycans, glycolipids, *O*-GlcNAc, and glycosaminoglycans. *N*-linked glycosylation occurs when glycans are attached to asparagine residues on the protein. *O*-linked glycans are most commonly attached to serine or threonine residues through the *N*-Acetylgalactosamine residue. Removal of oligosaccharides from glycoproteins, termed deglycosylation, is often used to simplify analysis of the peptide and/or glycan portion of a glycoprotein. Detailed knowledge of the glycan structures helps to correlate them to their respective function. To do this, tools are required for highly sensitive analysis of glycan chains. Both chemical and enzymatic methods exist for removing oligosaccharides from glycoproteins. However, chemical methods such as β -elimination with mild alkali (1) or mild hydrazinolysis (2) can be harsh and results in the degradation of the protein; whereas enzymatic methods are much gentler and can provide complete sugar removal with no protein degradation.

ADVANTAGES

- Low cost
- Easy Reaction Setup
- Digestion with a combination of enzymes can yield structural information
- Can be used under native and denaturing conditions

Endoglycosidase Selection Chart

| | PROTEIN DEGLYCOSYLA- TION MIX II (#P6044) | O-GLYCOSIDASE (#P0733 & #E0540) | PNGASE A (#P0707) | PNGASE F (#P0704 & #P0705) | REMOVE-IT PNGASE F (#P0706) | RECOM- BINANT PNGASE F (#P0708 & #P0709) | RAPID PNGASE F (#P0710) | RAPID PNGASE F, (NON-REDUCING FORMAT) (#P0711) | ENDO H (#P0702) & ENDO H ₁ (#P0703) | ENDO S (#P0741) | ENDO D (#P0742) | ENDO F2 (#P0772) | ENDO F3 (#P0771) |
|---|--|---------------------------------------|----------------------|----------------------------------|-----------------------------------|--|----------------------------|---|---|--------------------|--------------------|---------------------|---------------------|
| Deglycosylation of glycoproteins (<i>N</i> - and <i>O</i> -glycans) | • | | | | | | | | | | | | |
| Removal of <i>O</i> -glycans | • | • | | | | | | | | | | | |
| Removal of <i>N</i> -glycans from glycoproteins | • | | • | • | • | • | • | • | • | • | • | • | • |
| Removal of high mannose and hybrid <i>N</i> -glycans (leaving a GlcNAc attached to Asn) | | | | | | | | | • | | | | |
| Optional removal of the enzyme from the reaction | | | | | • | | | | | • | • | • | • |
| Removal of paucimannose <i>N</i> -glycans (GlcNAc attached to Asn) | | | | | | • | | | • | | • | | |
| Removal of <i>N</i> -glycans from IgGs (leaving a GlcNAc attached to Asn) | | | | | | | | | • | • | | | |
| Analysis of therapeutic glycoproteins, compliance with regulatory agencies | | | | | | • | • | • | | | | | |
| High throughput <i>N</i> -glycan analysis of monoclonal anti- bodies, regulatory compliance | | | | | | | • | • | | | | | |
| Glycomics | | | | • | • | • | • | • | • | • | • | • | • |
| Proteomics | | | | • (only GF) | • | • (only GF) | • | • | • | • | • | • | • |
| Determine <i>N</i> -glycan sites | | | | | | | | | • | • | • | • | • |
| Removal of <i>N</i> -glycans from plant and insect glycoproteins | | | • | | | | | | | | | | |

GF = Glycerol Free

References

1. Kakehi, K. et al. (1994) *J. Chromatogr. A*. 680, 209–215.
2. Royle, L. et al. (2002) *Anal. Biochem.* 304, 70–90.



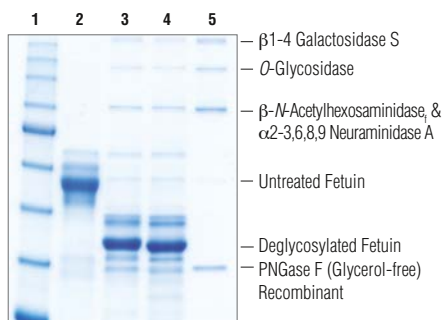
Protein Deglycosylation Mix II

Deglycosylation of a glycoprotein often requires more than one enzyme to completely remove all carbohydrate residues. PNGase F removes almost all *N*-linked oligosaccharides, while mono-saccharides on *O*-linked glycans must be removed by a series of exoglycosidases, such as β 1-4 Galactosidase S and β -*N*-Acetylhexosaminidase_r, until only the Gal β 1-3GalNAc (core 1) and/or the GlcNAc β 1-3GalNAc (core 3) cores remain attached to the core protein. *O*-Glycosidase can then remove these core structures leaving serine or threonine residues intact. Sialic acid residues, which will block the action of the *O*-Glycosidase, are easily removed by NEB's general α 2-3,6,8,9 Neuraminidase A.

The Protein Deglycosylation Mix II contains all of the enzymes, reagents, and controls needed to remove all *N*-linked and simple *O*-linked glycans as well as some complex *O*-linked glycans. This mix contains enzyme sufficient for 20 reactions or the cleavage of as much as 2 mg of glycoprotein. The Protein Deglycosylation Enzyme Mix II (100 μ l) is a single mix made up of five recombinant enzymes: PNGase F Glycerol Free Recombinant, *O*-Glycosidase, α 2-3,6,8,9 Neuraminidase A, β 1-4 Galactosidase S, and β -*N*-Acetylhexosaminidase_r. The mix is supplied with all of the reagents and controls required to complete the experiment under either non-denaturing (native) or reducing conditions. Components include 10X Deglycosylation Mix Buffer 1, 10X Deglycosylation Mix Buffer 2, and a Fetuin control containing sialylated *N*-linked and *O*-linked glycans. All of the enzymes and reagents included in the Protein Deglycosylation Mix II are Mass Spectrometry compatible. Following the deglycosylation reaction, samples are ready to be prepared for mass spectrometry analysis.

Protein Deglycosylation Mix IIP6044S

Enzymatic Deglycosylation of Bovine Fetuin



Enzymatic Deglycosylation of Bovine Fetuin under both native (10X Deglycosylation Mix Buffer 1) and reducing (10X Deglycosylation Mix Buffer 2) conditions. 20 μ g reactions were loaded onto a 10-20% Tris-glycine SDS-PAGE gel.

Lane 1: Color Prestained Protein Standard, Broad Range (11-245 kDa) (NEB #P7712)

Lane 2: 20 μ g untreated Fetuin control

Lane 3: 20 μ g Fetuin deglycosylated under native conditions with Deglycosylation Mix Buffer 1

Lane 4: 20 μ g Fetuin deglycosylated under reducing conditions with Deglycosylation Mix Buffer 2

Lane 5: 5 μ l Protein Deglycosylation Mix II



N-Linked Deglycosylation Enzymes

For structural analysis of asparagine-linked carbohydrates (*N*-linked glycans), sugars are released from the protein backbone by enzymes such as PNGase F, PNGase A, Endoglycosidase S, Endoglycosidase D and Endoglycosidase H.

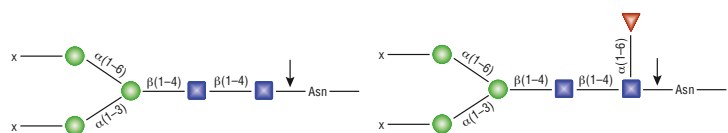
PNGase F (native and recombinant)

Peptide-*N*-Glycosidase F, also known as PNGase F, is an amidase which cleaves between the innermost GlcNAc and asparagine residues of high mannose, hybrid and complex oligosaccharides from *N*-linked glycoproteins. PNGase F from NEB is purified from *Flavobacterium meningosepticum*. A glycerol-free version of PNGase F is also offered for HPLC methods.

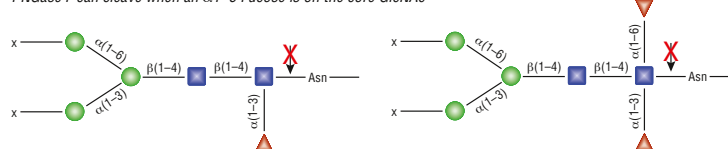
Detailed Specificity:

PNGase F hydrolyzes nearly all types of *N*-glycan chains from glycopeptides/proteins. PNGase F can cleave when an α 1–6 Fucose is on the core GlcNAc. PNGase F cannot cleave when an α 1–3 Fucose is on the core GlcNAc.

| | |
|---|----------|
| PNGase F | P0704S/L |
| PNGase F, Recombinant | P0708S/L |
| PNGase F (Glycerol-free) | P0705S/L |
| PNGase F (Glycerol-free), Recombinant | P0709S/L |



PNGase F hydrolyzes nearly all types of *N*-glycan chains from glycopeptides/proteins. [*x* = H or oligosaccharide].
PNGase F can cleave when an α 1–6 Fucose is on the core GlcNAc



PNGase F cannot cleave when an α 1–3 Fucose is on the core GlcNAc

References

1. Maley, F. et al. (1989) *Anal. Biochem.*, 180, 195–204.
2. Tretter, V. et al. (1991) *Eur. J. Biochem.* 199, 647–652.



Rapid PNGase F

A growing number of antibodies and antibody fusions are currently used as therapeutic agents. A conserved *N*-glycan at Asn297 of the Fc region of IgG is critical for functional activity. Moreover, some antibodies have additional *N*-glycans that, together with the conserved site, affect recognition, half-life, and immune reactions. Antibody glycosylation is heterogeneous, and variables in cell culture can increase glycan diversity. Monitoring glycosylation during production is essential to obtain the correct glycoprotein forms.

Obtaining an accurate *N*-glycan profile in the shortest time possible is essential for effective process control. Typically, enzymatic release of antibody *N*-glycans using PNGase F requires an incubation time of several hours, followed by glycan derivatization and analysis by liquid chromatography and/or mass spectrometry. In addition, incomplete deglycosylation can lead to biased results. Some glycans are easier to remove than others and unless deglycosylation is extensive, the profile obtained will not represent the correct composition of the therapeutic antibody.

ADVANTAGES

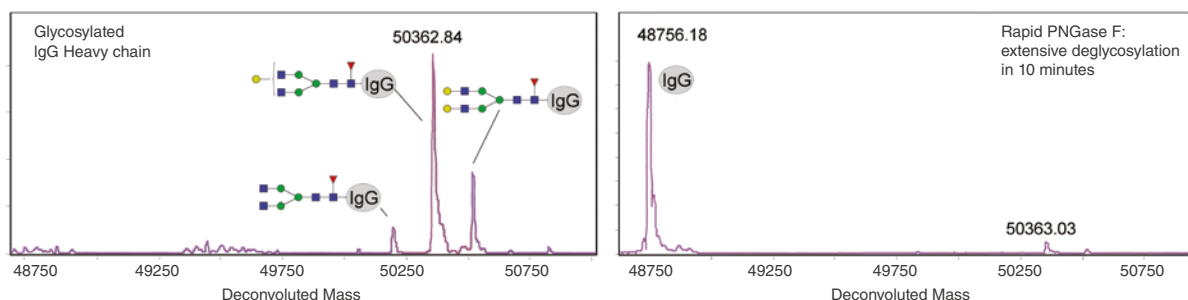
- Complete deglycosylation of antibodies and immunoglobulin fusion proteins in minutes
- Release of all *N*-glycans rapidly and without bias, ready for downstream chromatography or mass spectrometry analysis
- Optimal activity is ensured for 12 months
- Purified to > 99% homogeneity, as determined by SDS-PAGE
- Recombinant source

Rapid PNGase F

Deglycosylation in minutes for *N*-glycan analysis

Rapid PNGase F is an improved reagent that allows the complete and rapid deglycosylation of antibodies and immunoglobulin-fusion proteins in minutes. All *N*-glycans are released rapidly and without bias, and are ready to be prepared for downstream chromatography or mass spectrometry analysis. Rapid PNGase F creates an optimized workflow which reduces processing time without compromising sensitivity or reproducibility.

ESI-TOF analysis of an antibody before and after treatment with Rapid PNGase F



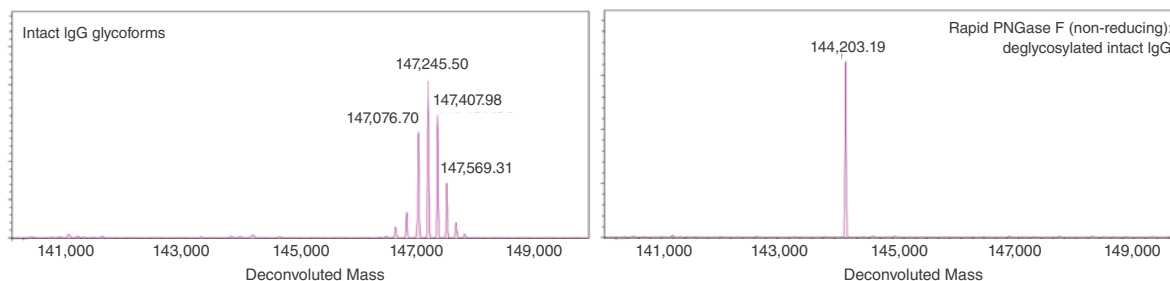
Rapid PNGase F (non-reducing format)

Deglycosylation in minutes for intact antibody analysis

Developed for proteomic applications, Rapid PNGase F (non-reducing format) is a reformulated version of Rapid PNGase F that allows the complete and rapid deglycosylation of antibodies and fusion proteins in minutes, while preserving disulfide bonds. All *N*-glycans are released rapidly and without bias, facilitating high throughput proteomics applications and methods for antibody characterization by mass spectrometry such as intact mass analysis. Rapid PNGase F (non-reducing format) combines the advantages of Rapid PNGase F (fast processing time), with the non-reducing conditions preserving quaternary structure.



ESI-TOF analysis of an antibody before and after treatment with Rapid PNGase F (non-reducing format)

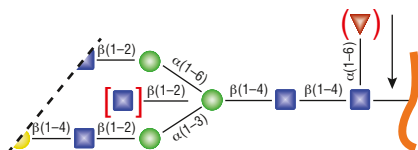


Detailed Specificity:

Rapid PNGase F & Rapid PNGase F (non-reducing format) cleave all complex, hybrid and high-mannose type glycans from antibodies and related proteins. Core α 1-3 fucosylation (found in immunoglobulins expressed in plant or insect cells) is resistant to all forms of PNGase F.

Rapid PNGase FP0710S

Rapid PNGase F (non-reducing format)P0711S



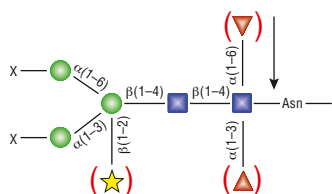
PNGase A

PNGase A is a recombinant amidase, which cleaves between the innermost GlcNAc and asparagine residues of high mannose, hybrid, and short complex oligosaccharides such as those found in plant and insect cells from N-linked glycoproteins and glycopeptides. PNGase A differs from PNGase F in that it cleaves N-linked glycans with or without α (1,3)-linked core fucose residues.

Detailed Specificity:

PNGase A hydrolyzes N-glycan chains from glycoproteins/peptides regardless of the presence of xylose or fucose. [x=H or Man or GlcNAc]

PNGase AP0707S/L



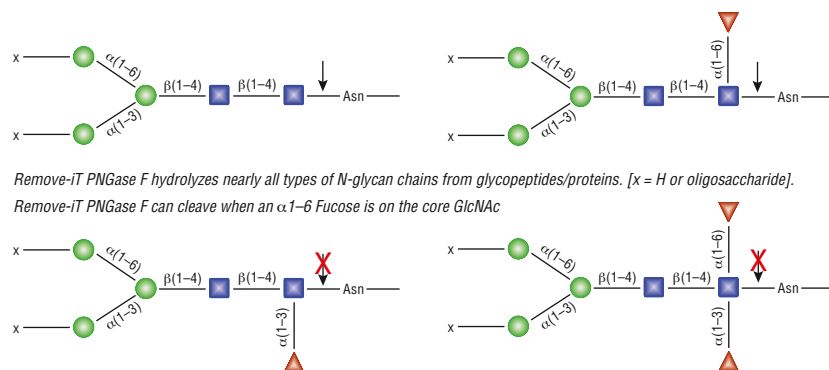
PNGase A hydrolyzes N-glycan chains from glycoproteins/peptides regardless of the presence of xylose or fucose. [x = H or Man or GlcNAc].



Remove-iT PNGase F

Remove-iT PNGase F is an amidase which cleaves between the innermost GlcNAc and asparagine residues of high mannose, hybrid, and complex oligosaccharides from *N*-linked glycoproteins. Remove-iT PNGase F is purified from *Flavobacterium meningosepticum* and is tagged with a chitin binding domain (CBD) for easy removal from a reaction. It is supplied glycerol free for optimal performance in HPLC and MS intensive methods..

Remove-iT PNGase F P0706S/L



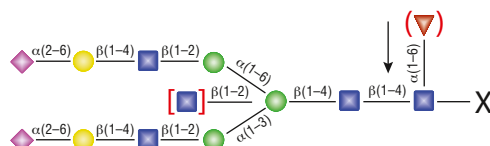
Remove-iT PNGase F cannot cleave when an α 1-3 Fucose is on the core GlcNAc

Endo S

Endo S is an endoglycosidase with a uniquely high specificity for removing *N*-linked glycans from the chitobiose core of the heavy chain of native IgG. Endo S is tagged with a chitin binding domain (CBD) for easy removal from a reaction and is supplied glycerol free for optimal performance in HPLC and MS intensive methods.

Endo S does not have a strict peptide requirement for activity, thus the “X” can be a protein, peptide, Asparagine, or free glycan. Endo S is active on a substrate with or without core α (1-6) fucosylation as well as with or without a bisecting *N*-acetylglucosamine. Triantennary and tetraantennary sialylated or asialo glycans are not a substrate for Endo S.

Endo S P0741S/L



ADVANTAGES

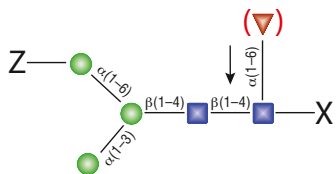
- Chitin Binding Domain (CBD) tag ensures easy removal from a reaction
- Fast reaction setup
- Compatible with protease inhibitor cocktails
- Glycerol-free formulation for optimal performance in HPLC and mass spec analysis



Endo D

Endo D also known as Endoglycosidase D is a recombinant glycosidase, which cleaves within the chitobiose core of paucimannose *N*-linked glycans, with or without extensions in the antennae. Endo D is active on both linear and branched upper arm extensions, and is useful for determining *N*-glycosylation sites. Endo D is tagged with a chitin binding domain (CBD) for easy removal from a reaction, and is supplied glycerol-free for optimal performance in HPLC and MS intensive methods.

Endo D..... P0742S/L

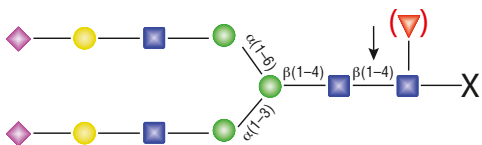


Endo D does not have a strict peptide requirement for activity, thus the "X" can be a protein, peptide or Asparagine. The upper branch, "Z", can be a "H", monosaccharide or oligosaccharide.

Endo F2

Endo F2 is a highly specific recombinant endoglycosidase which cleaves within the chitobiose core of asparagine-linked complex biantennary and high mannose oligosaccharides from glycoproteins and glycopeptides. Endo F2 cleaves biantennary glycans at a rate approximately 20 times greater than high mannose glycans. The activity of Endo F2 is identical on biantennary structures with and without core fucosylation. However, Endo F2 is not active on hybrid or tri- and tetra-antennary oligosaccharides. Endo F2 is tagged with a chitin binding domain (CBD) for easy removal from a reaction and is supplied glycerol free for optimal performance in HPLC and MS intensive methods.

Endo F2..... P0772S/L

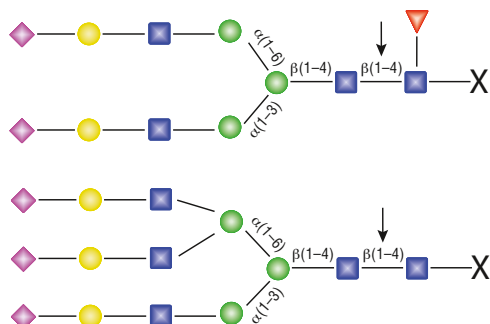




Endo F3

Endo F3 is a highly specific recombinant endoglycosidase which cleaves within the chitobiose core of asparagine-linked fucosylated-biantennary and triantennary complex oligosaccharides from glycoproteins. Endo F3 is tagged with a chitin binding domain (CBD) for easy removal from a reaction and is supplied glycerol free for optimal performance in HPLC and MS intensive methods.

Endo F3 P0771S/L

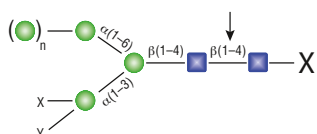


Endoglycosidase H

Endoglycosidase H (Endo H) is a recombinant glycosidase which cleaves within the chitobiose core of high mannose and some hybrid oligosaccharides from N-linked glycoproteins. Endo H_f is a recombinant protein fusion of Endoglycosidase H and maltose binding protein. It has identical activity to Endo H. Endo H and Endo H_f from NEB are cloned from *Streptomyces plicatus* and overexpressed in *E. coli*.

Endoglycosidase H P0702S/L

Endoglycosidase H_f P0703S/L



Endo H and Endo H_f cleave only high mannose structures
(n = 2–150, x = (Man)_x, y = H) and hybrid structures (n = 2,
x and/or y = AcNeu-Gal-GlcNAc) "X" can be a protein, peptide
or Asparagine.



O-Linked Deglycosylation Enzymes

For structural analysis of serine or threonine-linked carbohydrates (O-linked glycans), sugars are released from the protein backbone by either chemical or enzymatic methods. Removing O-linked glycan chains while rendering a protein intact for further examination can be a difficult task. Chemical methods, such as β -elimination, may result in incomplete sugar removal and degradation of the protein. On the other hand, enzymatic removal of O-linked glycans must be performed as a series of exoglycosidase digestions until only the Gal β 1-3GalNAc (core 1) and/or the GlcNAc β 1-3GalNAc (core 3) cores remains attached to the serine or threonine residue. NEB's *Enterococcus faecalis* O-Glycosidase, also known as Endo- α -N-Acetylgalactosaminidase, catalyzes the removal of core 1 and core 3 disaccharide structures with no modification of the serine or threonine residues (1). Any modification of the core structures, including sialylation, will block the action of the O-Glycosidase. Sialic acid residues are easily removed by a general Neuraminidase. In addition, exoglycosidases such as β 1-4 Galactosidase and β -N-Acetylglucosaminidase can be included in deglycosylation reactions to remove other complex modifications often known to be present on the core structures.

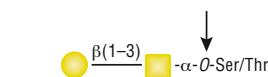
O-Glycosidase

O-Glycosidase, also known as Endo- α -N-Acetylgalactosaminidase, catalyzes the removal of Core 1 and Core 3 O-linked disaccharides from glycoproteins. O-Glycosidase from NEB is cloned from *Enterococcus faecalis* and expressed in *E. coli*.

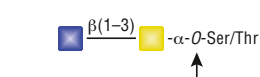
| | |
|---|----------|
| O-Glycosidase (Endo- α -N-Acetylgalactosaminidase) | P0733S/L |
| α 2-3,6,8 Neuraminidase | P0720S/L |
| O-Glycosidase & Neuraminidase Bundle | E0540S |

Reference

1. Koutsoulis, D., Landry, D. and Guthrie, E.P. (2008) *Glycobiology* 18, 799–805.



A. 5 mg substrate, 1 hr incubation, Core 1 Cleavage



B. 5 mg substrate, 1 hr incubation, Core 3 Cleavage

Companion Products

Endoglycosidase Reaction Buffer Pack

The Endoglycosidase Reaction Buffer Pack contains 1 ml of every buffer necessary for optimal activity of a deglycosylation reaction including 10X GlycoBuffer 2, 10X GlycoBuffer 3, 10X Glycoprotein Denaturing Buffer and 10% NP-40.

Endoglycosidase Buffer Pack..... B0701S

RNase B

RNase B is a high mannose glycoprotein that can be used as a positive control for endoglycosidases that cleave N-linked carbohydrates. RNase B has a single N-linked glycosylation site which makes it ideal for SDS-PAGE gel shift assays. It has an intact molecular weight of 17,000 daltons, and a molecular weight of 13,683 daltons after deglycosylation.

RNase B..... P7817S

Fetuin

Fetuin is a glycoprotein containing sialylated N-linked and O-linked glycans that can be used as a positive control for endoglycosidase enzymes.

Fetuin..... P6042S

Rapid PNGase F Antibody Standard

Rapid PNGase F Antibody Standard is a murine anti-MBP monoclonal antibody, isotype IgG2a.

It is comprised of two heavy chains which are each approximately 49 kDa, as well as two light chains which are each approximately 24.4 kDa. This antibody standard can be used as a positive control for Rapid PNGase F.

Rapid PNGase F Antibody Standard..... P6043S



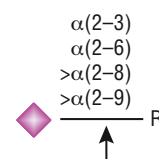
Exoglycosidase Enzymes

NEB offers a wide selection of exoglycosidases for glycobiology research. Exoglycosidases cleave a monosaccharide from the non-reducing end of an internal glycosidic linkage in an oligosaccharide or polysaccharide. Many of these reagents are recombinant, and all undergo several quality control assays, enabling us to provide products with lower unit cost, high purity, and reduced lot-to-lot variation. All of our glycosidases are tested for contaminants. Since *p*-nitrophenyl-glycosides are not hydrolyzed by some exoglycosidases, we use only fluorescently-labeled oligosaccharides to assay activity and screen for contaminating glycosidases.

α 2-3,6,8,9 Neuraminidase A

Neuraminidase is the common name for Acetyl-neuraminyl hydrolase (Sialidase). α 2-3,6,8,9 Neuraminidase A, cloned from *Arthrobacter ureafaciens*, catalyzes the hydrolysis of all linear and branched non-reducing terminal sialic acid residues from glycoproteins and oligosaccharides. The enzyme releases α 2-3 and α 2-6 linkages at a slightly higher rate than α 2-8 and α 2-9 linkages.

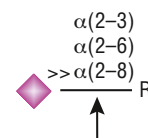
α 2-3,6,8,9 Neuraminidase A P0722S/L



α 2-3,6,8 Neuraminidase

α 2-3,6,8 Neuraminidase, cloned from *Clostridium perfringens*, catalyzes the hydrolysis of α 2-3, α 2-6 and α 2-8 linked *N*-acetylneuraminic acid residues from glycoproteins and oligosaccharides.

α 2-3,6,8 Neuraminidase P0720S/L

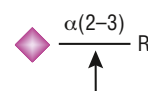


α 2-3 Neuraminidase S

α 2-3 Neuraminidase S is a highly specific exoglycosidase that catalyzes the hydrolysis of α 2-3 linked *N*-acetylneuraminic acid residues from glycoproteins and oligosaccharides.

α 2-3 Neuraminidase S is cloned from *Streptococcus pneumoniae* and expressed in *E. coli*.

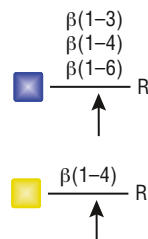
α 2-3 Neuraminidase S P0743S/L



β -N-Acetylhexosaminidase_f

β -N-Acetylhexosaminidase_f, cloned from *Streptomyces plicatus*, is a recombinant protein fusion of β -N-Acetylhexosaminidase and maltose binding protein with identical activity to β -N-Acetylhexosaminidase. It catalyzes the hydrolysis of terminal β -N-Acetylgalactosamine and glucosamine residues from linear oligosaccharides.

β -N-Acetylhexosaminidase_f P0721S/L



References

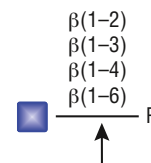
I. Iwamori, M., Ohta, Y., Uchida, Y. and Tsukada, Y. (1997) *Glycocon. J.*, 1, 67-73.



β -N-Acetylglucosaminidase S

β -N-Acetylglucosaminidase S, cloned from *Streptococcus pneumoniae*, is a highly specific exoglycosidase that catalyzes the hydrolysis of terminal, non-reducing β -N-Acetylglucosamine residues from oligosaccharides. β -N-Acetylglucosaminidase S is able to efficiently cleave bisecting β -N-Acetylglucosamine residues.

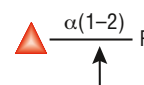
β -N-Acetylglucosaminidase S P0744S/L



α 1-2 Fucosidase

α 1-2 Fucosidase, cloned from *Xanthomonas manihotis*, is a highly specific exoglycosidase that catalyzes the hydrolysis of linear α 1-2 linked fucose residues from oligosaccharides. A linear substrate is defined as having no branching on the adjacent residue.

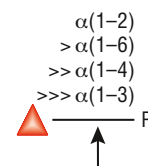
α 1-2 Fucosidase P0724S/L



α 1-2,3,4,6 Fucosidase

α 1-2,3,4,6 Fucosidase, cloned from bovine kidney, is a broad specificity exoglycosidase that catalyzes the hydrolysis of α 1-2, α 1-3, α 1-4 and α 1-6 linked fucose residues from oligosaccharides. α 1-2,3,4,6 Fucosidase cleaves α 1-2 and α 1-6 fucose residues more efficiently than other linkages, and has slight activity towards α 1-3 fucose residues.

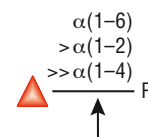
α 1-2,3,4,6 Fucosidase P0748S/L



α 1-2,4,6 Fucosidase O

α 1-2,4,6 Fucosidase O, cloned from *Omnitrophica*, is a broad specificity exoglycosidase that catalyzes the hydrolysis of terminal α 1-2, α 1-4 and α 1-6 linked fucose residues from oligosaccharides and glycoproteins. α 1-2,4,6 Fucosidase O cleaves α 1-6 fucose residue more efficiently than other linkages.

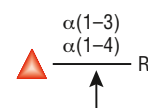
α 1-2,4,6 Fucosidase O P0749S/L



α 1-3,4 Fucosidase

α 1-3,4 Fucosidase, (also known as AMF) is a broad specificity exoglycosidase that catalyzes the hydrolysis of α 1-3 and α 1-4 linked fucose residues from oligosaccharides and glycoproteins.

α 1-3,4 Fucosidase P0769S/L

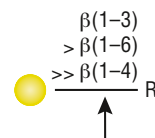




β 1-3 Galactosidase

β 1-3 Galactosidase, cloned from *Xanthomonas manihotis*, is a highly specific exoglycosidase that catalyzes the hydrolysis of β 1-3 and, at a much lower rate, β 1-6 linked galactose residues from oligosaccharides. The approximate kinetic data show >100-fold preference for β 1-3 over β 1-6 linkages and >500-fold preference for β 1-3 over β 1-4 linkages.

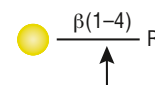
β 1-3 Galactosidase P0726S/L



β 1-4 Galactosidase S

β 1-4 Galactosidase S, cloned from *Streptococcus pneumoniae*, is a highly specific exoglycosidase that catalyzes the hydrolysis of β 1-4 linked galactose residues from oligosaccharides.

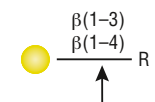
β 1-4 Galactosidase S P0745S/L



β 1-3,4 Galactosidase

β 1-3,4 Galactosidase, cloned from bovine testis and also known as BTG, is a highly specific exoglycosidase that catalyzes the hydrolysis of terminal β 1-3 and β 1-4 linked galactose residues from oligosaccharides.

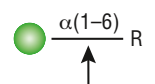
β 1-3,4 Galactosidase P0746S/L



α 1-6 Mannosidase

α 1-6 Mannosidase, cloned from *Xanthomonas manihotis*, is a highly specific exoglycosidase that removes unbranched α 1-6 linked mannose residues from oligosaccharides. When used in conjunction with α 1-2,3 Mannosidase, the α 1-6 Mannosidase will cleave α 1-6 mannose residues from branched carbohydrate substrates.

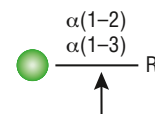
α 1-6 Mannosidase P0727S/L



α 1-2,3 Mannosidase

α 1-2,3 Mannosidase, cloned from *Xanthomonas manihotis*, is a highly specific exoglycosidase that catalyzes the hydrolysis of α 1-2 and α 1-3 linked mannose residues from oligosaccharides.

α 1-2,3 Mannosidase P0729S/L

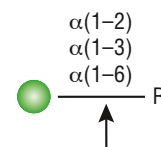




α 1-2,3,6 Mannosidase

α 1-2,3,6 Mannosidase, cloned from Jack Bean, and also known as JBM, is a broad specificity exoglycosidase that catalyzes the hydrolysis of terminal α 1-2, α 1-3 and α 1-6 linked mannose residues from oligosaccharides. α 1-2,3,6 Mannosidase has a slight preference for α 1-2 mannose residues over α 1-3 and α 1-6 mannose residues.

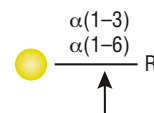
α 1-2,3,6 Mannosidase P0768S/L



α 1-3,6 Galactosidase

α 1-3,6 Galactosidase, cloned from *Xanthomonas manihotis*, is a highly specific exoglycosidase that catalyzes the hydrolysis of α 1-3 and α 1-6 linked galactose residues from oligosaccharides.

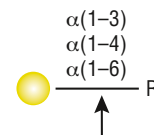
α 1-3,6 Galactosidase P0731S/L



α 1-3,4,6 Galactosidase

α 1-3,4,6 Galactosidase, cloned from green coffee bean, is a highly specific exoglycosidase that catalyzes the hydrolysis of α 1-3, α 1-4 and α 1-6 linked galactose residues from oligosaccharides.

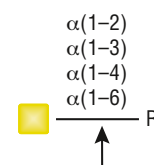
α 1-3,4,6 Galactosidase P0747S/L



α -N-Acetylgalactosaminidase

α -N-Acetylgalactosaminidase, cloned from *Chryseobacterium meningosepticum*, is a highly specific exoglycosidase that catalyzes the hydrolysis of α -N-acetylgalactosamine residues from oligosaccharides and N-linked glycans attached to proteins.

α -N-Acetylgalactosaminidase P0734S/L



GlycoBuffer Compositions

1X GlycoBuffer 1:

50 mM sodium acetate (pH 5.5 @ 25°C), 5 mM CaCl₂

1X GlycoBuffer 2:

50 mM sodium phosphate (pH 7.5 @ 25°C)

1X GlycoBuffer 3:

50 mM sodium acetate (pH 6.0 @ 25°C)

1X GlycoBuffer 4:

50 mM sodium acetate (pH 4.5 @ 25°C)



Heparin Lyase Enzymes

Heparin Lyase enzymes, also called Heparinases, are enzymes that cleave the glycosidic linkage between hexosamines and uronic acids and are known to cleave heparin and heparin sulfate (HS) chains selectively, via an elimination mechanism. Heparinase enzymes create a double bond on the non-reducing end of the uronic acid that absorbs at 232 nm and can be used for the detection of oligosaccharide and disaccharide products. Three Heparinase enzymes are available: *Bacteroides* Heparinase I, Heparinase II and Heparinase III. Heparinase I cleaves highly sulfated heparin/HS chains, Heparinase III cleaves less sulfated HS chains, while Heparinase II cleaves domains of both high and low sulfation on both heparin and HS. Heparinase I, II and III used in combination can produce a near-complete depolymerization of heparin/HS polysaccharide chains to disaccharides.

Bacteroides Heparinase I

Bacteroides Heparinase I cloned from *Bacteroides eggerthii*, also called Heparin Lyase I, is active on heparin and the highly sulfated domains of heparan sulfate. The reaction yields oligosaccharide products containing unsaturated uronic acids which can be detected by UV spectroscopy at 232 nm.

Detailed Specificity:

Bacteroides Heparinase I cleaves between *N*-sulfated hexosamines and 2-*O*-sulfated iduronic acid residues as well as the 2-*O*-sulfated glucuronic acid residue. The 2-*O*-sulfated uronic acid residue is essential for the activity of *Bacteroides* Heparinase I and 6-*O*-sulfation of GlcNSO₃ does not hinder enzyme activity. Limited digest of porcine mucosal heparin with the *Bacteroides* Heparinase I results in heparin oligosaccharides with a lower extent of sulfation as reported (1).

Bacteroides Heparinase I P0735S/L

Bacteroides Heparinase II

Bacteroides Heparinase II cloned from *Bacteroides eggerthii*, also called Heparin Lyase II, is a low specificity enzyme that is active on both heparin and heparan sulfate. The reaction yields oligosaccharide products containing unsaturated uronic acids which can be detected by UV spectroscopy at 232 nm.

Detailed Specificity:

Bacteroides Heparinase II cleaves the glycosidic bond between *N*-sulfated glucuronic or iduronic acid residues. When used alone this enzyme rarely yields complete depolymerization of a polysaccharide chain, however disaccharide analysis is enhanced when used in combination with Heparinase I and III.

Bacteroides Heparinase II P0736S/L

Bacteroides Heparinase III

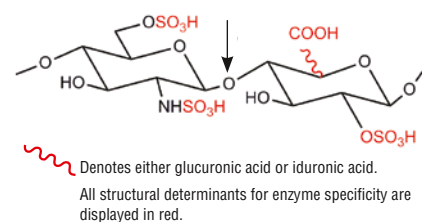
Bacteroides Heparinase III cloned from *Bacteroides eggerthii*, also called Heparin Lyase III, is active on both heparin and heparan sulfate. The reaction yields oligosaccharide products containing unsaturated uronic acids which can be detected by UV spectroscopy at 232 nm.

Detailed Specificity:

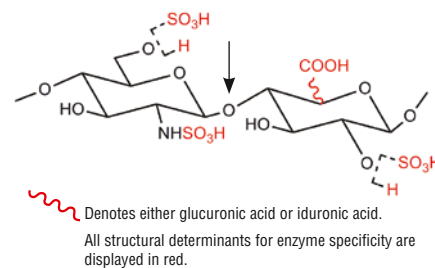
Bacteroides Heparinase III can cleave the glycosidic bond between hexosamines and either iduronic acid or glucuronic acid residues, and is active in the presence of 6-sulfation.

Bacteroides Heparinase III P0737S/L

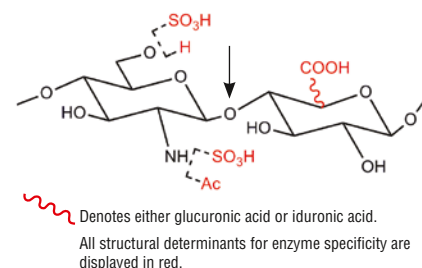
Note: Heparin Hexasaccharide MS Standard 6 (NEB #P0738S) and Heparin Hexasaccharide MS Standard 7 (NEB #P0739S) are available by special order. Contact info@neb.com for details.



Bacteroides Heparinase I specificity.



Bacteroides Heparinase II specificity.



Bacteroides Heparinase III specificity.

References

1. Merchant, Z.M., Kim, Y.S., Rice, K.G., Linhardt, R. (1985) *J. Biol. Chem.* 229, 369–377.



Glycoproteomics

NEB offers a selection of high purity proteases and standards for use in concert with our glycosidases for proteomic and glycoproteomic applications. Our proteases are of the highest quality, show no lot-to-lot variation and are offered at exceptional value. They are tested to assure a lack of contaminating proteins as well as high activity. They are used daily at NEB for internal research projects with a wide range of state-of-the-art mass spectrometers.

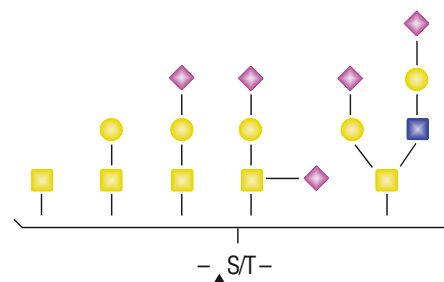
O-Glycoprotease

O-Glycoprotease is a highly specific protease that cleaves the peptide bonds of a glycoprotein or glycopeptide immediately N-terminal to a serine or threonine residue containing a mucin-type O-linked glycan with or without sialylation. O-Glycoprotease has a 6xHis-tag for easy removal from a reaction using nickel affinity resins.

O-GlycoproteaseP0761S

ADVANTAGES

- Enables O-glycosylation site mapping and O-glycan structural determination
- Efficiently cleaves glycoproteins with or without sialic acid, no neuraminidase treatment necessary
- Recombinant enzyme with no detectable non-specific protease contaminating activities
- Can be used under native or denaturing conditions
- Glycerol-free for optimal performance in HPLC and mass spectrometry analysis



IdeZ Protease (IgG-specific)

IdeZ Protease (IgG-specific) is a recombinant antibody specific protease cloned from *Streptococcus equi* subspecies *zooepidemicus* that recognizes all human, sheep, monkey, and rabbit IgG subclasses, specifically cleaving at a single recognition site below the hinge region, yielding a homogenous pool of F(ab')₂ and Fc fragments. IdeZ Protease has significantly improved activity against murine IgG2a and IgG3 subclasses compared to IdeS Protease.

Detailed Specificity:

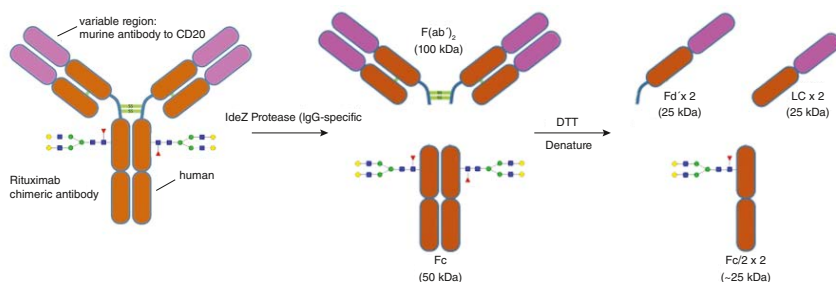
human IgG1, IgG3, IgG4: CPAPELLG[▼]GPSVF

human IgG2: CPAPPVA[▼]GPSVF

murine IgG2a: CPAPNLLG[▼]GPSVF

murine IgG3: CPPGNILG[▼]GPSVF

IdeZ Protease (IgG-specific)P0770S



Digestion of IgG with IdeZ Protease (IgG-specific), followed by denaturation.

Notes: IdeZ Protease efficiently cleaves human, humanized, chimeric, sheet, rabbit and monkey IgG as well as mouse IgG2a and IgG3. IdeZ Protease will also cleave Fc-fusion proteins, such as Enbrel.

IdeZ Protease does not cleave mouse IgG1 or IgG2b, rat, porcine, bovine or goat IgG. It also does not cleave non-IgG isotypes including IgA, IgM, IgD and IgE.

Gal ● Glc ● Man ● GalNAc ■ GlcNAc ■ Fuc ▲ NeuAc ◆ R = any sugar



Trypsin-digested BSA MS Standard (CAM-modified)

A complex mixture of peptides produced by the tryptic digest of Bovine Serum Albumin (BSA) that has been reduced and alkylated with iodacetamide (CAM modified). This peptide mixture is free of salts, glycerol and detergents and can therefore be used to standardize Matrix-Assisted Laser Desorption/Ionization Time-Of-Flight (MALDI-TOF) or Electrospray Ionization (ESI) mass spectrometers (TOF, Q-TOF, Ion Trap, or Orbitrap) using a standardization range of 500-2400 Da.

Trypsin-digested BSA MS Standard (CAM-modified)P8108S

Trypsin-ultra,TM Mass Spectrometry Grade

Trypsin-ultra, Mass Spectrometry Grade is a serine endopeptidase isolated from Bovine pancreas. Trypsin is the most widely used enzyme in proteomics. It selectively cleaves peptide bonds C-terminal to lysine and arginine residues. Trypsin-ultra is treated with L-(tosylamido-2-phenyl) ethyl chloromethyl ketone (TPCK) to inactivate any remaining chymotryptic activity. It is modified by acetylation of the ε-amino groups of lysine residues to prevent autolysis. Trypsin-ultra cleaves at Lys-Pro and Arg-Pro bonds at a much slower rate than when Lys and Arg are N-terminal to other residues.

Xxx-Arg▼Xxx

Xxx-Lys▼Xxx

Trypsin-ultra, Mass Spectrometry GradeP8101S

α-Lytic Protease

α-Lytic Protease (aLP) cleaves after Threonine (T), Alanine (A), Serine (S) and Valine (V) residues. Its specificity makes it an orthogonal and alternative protease to others commonly used in proteomics applications, including trypsin and chymotrypsin. Peptides generated by aLP are of similar average length to those of Trypsin.

XX-T/A/S/V▼XX

α-Lytic Protease P8113S/L

Endoproteinase LysC

LysC is a serine endoproteinase, isolated from *Lysobacter enzymogenes*, that cleaves peptide bonds at the carboxyl side of lysine. LysC is a sequencing grade enzyme and is suitable for proteomics and glycobiology applications.

XX-Lys▼XXX

Endoproteinase LysCP8109S

Endoproteinase AspN

Endoproteinase AspN (flavastacin) is a zinc metalloendopeptidase which selectively cleaves peptide bonds N-terminal to aspartic acid residues.

XX▼Asp-XXX

Endoproteinase AspNP8104S

Endoproteinase GluC

Endoproteinase GluC (*Staphylococcus aureus* Protease V8) is a serine proteinase which preferentially cleaves peptide bonds C-terminal to glutamic acid residues. Endoproteinase GluC also cleaves at aspartic acid residues at a rate 100-300 times slower than at glutamic acid residues. *Staphylococcus aureus* Protease V8 gene was cloned and expressed in *Bacillus subtilis*. Endoproteinase GluC is observed as a single band on SDS-PAGE and TOF-MS and is intended for use in mass spectrometry and protein sequencing.

Xxx-Glu▼Xxx

Xxx-Asp▼Xxx

Endoproteinase GluCP8100S



Ordering Information

Deglycosylation Enzymes

| PRODUCT | NEB # | SIZE |
|---------------------------------------|----------|---|
| Protein Deglycosylation Mix II | P6044S | 20 reactions |
| PNGase A | P0707S/L | 150/750 units (5,000 units/ml) |
| Rapid PNGase F | P0710S | 50 reactions |
| Rapid PNGase F (non-reducing format) | P0711S | 50 reactions |
| PNGase F | P0704S/L | 15,000/75,000 units (500,000 units/ml) |
| PNGase F, Recombinant | P0708S/L | 15,000/75,000 units (500,000 units/ml) |
| PNGase F (Glycerol-free) | P0705S/L | 15,000/75,000 units (500,000 units/ml) |
| PNGase F (Glycerol-free), Recombinant | P0709S/L | 15,000/75,000 units (500,000 units/ml) |
| Remove-iT PNGase F | P0706S/L | 6,750/33,750 units (225,000 units/ml) |
| Endo F2 | P0772S | 480 units (8,000 units/ml) |
| Endo F3 | P0771S | 240 units (8,000 units/ml) |
| Endo S | P0741S/L | 6,000/30,000 units (200,000 units/ml) |
| Endo D | P0742S/L | 1,500/7,500 units (50,000 units/ml) |
| Endo H | P0702S/L | 10,000/50,000 units (500,000 units/ml) |
| Endo H ₁ | P0703S/L | 100,000/500,000 units (1,000,000 units/ml) |
| O-Glycosidase | P0733S/L | 2,000,000/10,000,000 units (40,000,000 units/ml) |
| O-Glycosidase & Neuraminidase Bundle | E0540S | 2,000,000 units O-Glycosidase & 2,000 units Neuraminidase |

Heparin Lyases

| PRODUCT | NEB # | SIZE |
|----------------------------|----------|---------------------------------------|
| Bacteroides Heparinase I | P0735S/L | 240 units/600 units (12,000 units/ml) |
| Bacteroides Heparinase II | P0736S/L | 80 units/200 units (4,000 units/ml) |
| Bacteroides Heparinase III | P0737S/L | 14 units/35 units (700 units/ml) |

Glycoproteomics

| PRODUCT | NEB # | SIZE |
|--|----------|---------------|
| O-Glycoprotease | P0761S | 200 reactions |
| IdeZ Protease (IgG-specific) | P0770S | 4,000 units |
| Trypsin-digested BSA MS Standards (CAM Modified) | P8108S | 500 pmol |
| Trypsin-ultra, Mass Spectrometry Grade | P8101S | 100 µg |
| α-Lytic Protease | P8113S/L | 20/100 µg |
| Endoproteinase LysC | P8109S | 20 µg |
| Endoproteinase AspN | P8104S | 50 µg |
| Endoproteinase GluC | P8100S | 50 µg |

Exoglycosidase Enzymes

| PRODUCT | NEB # | SIZE |
|---------------------------------------|----------|--------------------------------------|
| α-N-Acetylgalactosaminidase | P0734S/L | 3,000/15,000 units (20,000 units/ml) |
| β-N-Acetylglucosaminidase S | P0744S/L | 100/500 units (4,000 units/ml) |
| β-N-Acetylhexosaminidase ₁ | P0721S/L | 500/2,500 units (5,000 units/ml) |
| α-1-2 Fucosidase | P0724S/L | 1,000/5,000 units (20,000 units/ml) |
| α-1-2,3,4,6 Fucosidase | P0748S/L | 400/2,000 units (8,000 units/ml) |
| α-1-2,4,6 Fucosidase O | P0749S/L | 80/400 units (2,000 units/ml) |
| α-1-3,4 Fucosidase | P0769S/L | 200/1,000 units (4,000 units/ml) |
| α-1-3,6 Galactosidase | P0731S/L | 100/500 units (4,000 units/ml) |
| α-1-3,4,6 Galactosidase | P0747S/L | 200/1,000 units (8,000 units/ml) |
| β-1-3 Galactosidase | P0726S/L | 500/2,500 units (10,000 units/ml) |
| β-1-3,4 Galactosidase | P0746S/L | 400/2,000 units (8,000 units/ml) |
| β-1-4 Galactosidase S | P0745S/L | 400/2,000 units (8,000 units/ml) |
| α-1-2,3 Mannosidase | P0729S/L | 640/3,200 units (32,000 units/ml) |
| α-1-2,3,6 Mannosidase | P0768S/L | 80/400 units (2,000 units/ml) |
| α-1-6 Mannosidase | P0727S/L | 800/4,000 units (40,000 units/ml) |
| α-2-3,6,8 Neuraminidase | P0720S/L | 2,000/10,000 units (50,000 units/ml) |
| α-2-3,6,8,9 Neuraminidase A | P0722S/L | 800/4,000 units (20,000 units/ml) |
| α-2-3 Neuraminidase S | P0743S/L | 400/2,000 units (8,000 units/ml) |

Companion Products

| PRODUCT | NEB # | SIZE |
|---|----------|-------------------|
| Endoglycosidase Reaction Buffer Pack | B0701S | 4 x 1 ml |
| Fetuin | P6042S | 500 µg (10 mg/ml) |
| RNase B | P7817S | 250 µg (5 mg/ml) |
| Rapid PNGase F Antibody Standard | P6043S | 250 µg (5 mg/ml) |
| Chitin Magnetic Beads | E8036S/L | 5/25 ml |
| 6-Tube Magnetic Separation Rack | S1506S | 6 tubes |
| 50 ml Magnetic Separation Rack | S1507S | 4 tubes |
| 12-Tube Magnetic Separation Rack | S1509S | 12 tubes |
| 2-Tube Magnetic Separation Rack | S1510S | 2 tubes |
| 96-Well Microtiter Plate Magnetic Separation Rack | S1511S | 96-well |

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